

Functional curation of the *Sulfolobus solfataricus* P2 and *S. acidocaldarius* 98-3 complete genome sequences

Domink Esser · Theresa Kouril · Melanie Zaparty · Pawel Sierocinski · Patricia P. Chan · Todd Lowe · John Van der Oost · Sonja-Verena Albers · Dietmar Schomburg · Kira S. Makarova · Bettina Siebers

Received: 27 July 2011 / Accepted: 28 July 2011 / Published online: 13 September 2011
© Springer 2011

Abstract The thermoacidophiles *Sulfolobus solfataricus* P2 and *S. acidocaldarius* 98-3 are considered key model organisms representing a major phylum of the Crenarchaeota. Because maintaining current, accurate genome information is indispensable for modern biology, we have updated gene function annotation using the arCOGs database, plus other available functional, structural and

phylogenetic information. The goal of this initiative is continuous improvement of genome annotation with the support of the *Sulfolobus* research community.

Keywords Archaea · Thermoacidophiles · Genome analysis · Genomics

Communicated by G. Antranikian.

D. Esser · T. Kouril · B. Siebers (✉)
Molecular Enzyme Technology and Biochemistry, Biofilm Centre, Faculty of Chemistry, University of Duisburg-Essen, Universitätsstraße 5, 45141 Essen, Germany
e-mail: bettina.siebers@uni-due.de

M. Zaparty
Institute for Molecular and Cellular Anatomy, University of Regensburg, Universitätsstr. 31, 93053 Regensburg, Germany

P. Sierocinski · J. Van der Oost
Laboratory of Microbiology, Wageningen University, Dreijenplein 10, 6703 HB Wageningen, The Netherlands

P. P. Chan · T. Lowe
Department of Biomolecular Engineering, University of California Santa Cruz, Santa Cruz, CA, USA

S.-V. Albers
Molecular Biology of Archaea, Max Planck Institute for Terrestrial Microbiology, Karl-von-Frisch Strasse, 35043 Marburg, Germany

D. Schomburg
Department of Bioinformatics and Biochemistry, Technical University Braunschweig, Langer Kamp 19b, 38106 Braunschweig, Germany

K. S. Makarova
National Center for Biotechnology Information, National Institutes of Health, Bethesda, MD 20894, USA

Both *Sulfolobus* species are obligate aerobes that thrive in thermoacidophilic habitats. *S. solfataricus* (strain P2, DSM1617; Zillig et al. 1980) optimally grows at 80°C and pH 3.2/3.5; *S. acidocaldarius* (98-3, DSM 639; Brock et al. 1972) has similar optimal growth between 75 and 80°C and pH 2–3. Whereas *S. solfataricus* grows heterotrophically on peptidolytic constituents and a wide variety of simple and complex sugars, *S. acidocaldarius* relies on tryptone for heterotrophic growth, but has enhanced growth with the addition of selected carbohydrates, such as sucrose, xylose, and glucose (Grogan 1989; Zaparty and Siebers 2010).

The genome of the *S. solfataricus* strain P2 is nearly 3 million base pairs (Mbp) with 3,033 predicted open reading frames (ORFs) (She et al. 2001b). In particular, this genome exhibits a high level of plasticity due in part to 200 diverse insertion sequence elements (Martusewitsch et al. 2000; Redder et al. 2001; She et al. 2001a; Brügger et al. 2002, 2004). In contrast, the genome of *S. acidocaldarius* is significantly more compact at 2.2 Mbp with 2,329 predicted ORFs, notably lacking insertion sequence elements (Chen et al. 2005).

S. solfataricus and *S. acidocaldarius* have become model organisms because they are easily maintained in the laboratory under defined growth conditions, and have recently developed gene expression and deletion systems (Worthington et al. 2003; Albers et al. 2006; Deng et al. 2009; Wagner et al. 2009). With complete genome

sequences, both organisms offer exciting opportunities for metabolic engineering, synthetic biology, and white biotechnology (Frazzetto 2003).

In the *Sulfolobus* Systems Biology (SulfoSYS) project, the effect of temperature change on a biological network, i.e. central carbohydrate metabolism, has been studied using biochemistry as well as genome-based high-throughput approaches (Albers et al. 2009; Pham et al. 2009; Zaparty et al. 2009). To provide a solid basis for future post-genome analyses, both *Sulfolobus* genomes have been updated using the recently developed clusters of orthologous groups database that is specific to archaeal genomes (arCOGs) (Makarova et al. 2007). This has enabled assignment of general or specific function to more than 770 protein-coding genes formerly annotated as hypothetical proteins.

Additional experimental, structural, and phylogenetic information were integrated from BRENDA (<http://www.brenda-enzymes.org/>), scientific literature searches (<http://www.ncbi.nlm.nih.gov/pubmed/>), and information from SulfoSYS partners. This information has allowed improvement and/or verification of annotation for 175 ORFs from *S. solfataricus* P2 (14 from orthologs reported in other *S. solfataricus* strains) and 33 ORFs from *S. acidocaldarius*. Recently published deep sequencing data for *S. solfataricus* transcription start sites (Wurtzel et al. 2010) has also enabled improved start codon positioning for 162 ORFs, as well as inclusion of 80 new ORFs.

The goal of this initiative is to continuously improve genome annotation with involvement of the scientific community. A submission form for newly characterized enzymes is available at the SulfoSYS homepage (<http://www.sulfosys.com/>). The curated *S. solfataricus* P2 and *S. acidocaldarius* 98-3 genomes are available at the SulfoSYS homepage (<http://www.sulfosys.com>, see “Hot Stuff”, Genome annotation) as well as the UCSC Archaeal Genome Browser (<http://archaea.ucsc.edu/sulfoSYS/>).

Acknowledgments The project has been performed in the course of the transnational SysMO initiative within the *Sulfolobus* Systems Biology “SulfoSYS” project. Partners gratefully acknowledge financial support from their respective national funding agencies (BMBF, NWO) as well as partner universities.

References

- Albers SV et al (2006) Production of recombinant and tagged proteins in the hyperthermophilic archaeon *Sulfolobus solfataricus*. *Appl Environ Microbiol* 72:102–111
- Albers SV et al (2009) SulfoSYS (*Sulfolobus* Systems Biology): towards a silicon cell model for the central carbohydrate

- metabolism of the archaeon *Sulfolobus solfataricus* under temperature variation. *Biochem Soc Trans* 37:58–64
- Brock TD, Brock KM, Belly RT, Weiss RL (1972) *Sulfolobus*: a new genus of sulfur-oxidizing bacteria living at low pH and high temperature. *Arch Mikrobiol* 84:54–68
- Brügger K, Redder P, She Q, Confalonieri F, Zivanovic Y, Garrett RA (2002) Mobile elements in archaeal genomes. *FEMS Microbiol Lett* 206:131–141
- Brügger K, Torarinsson E, Redder P, Chen L, Garrett RA (2004) Shuffling of *Sulfolobus* genomes by autonomous and non-autonomous mobile elements. *Biochem Soc Trans* 32:179–183
- Chen L et al (2005) The genome of *Sulfolobus acidocaldarius*, a model organism of the Crenarchaeota. *J Bacteriol* 187:4992–4999
- Deng L, Zhu H, Chen Z, Liang YX, She Q (2009) Unmarked gene deletion and host-vector system for the hyperthermophilic crenarchaeon *Sulfolobus islandicus*. *Extremophiles* 13:735–746
- Frazzetto G (2003) White biotechnology. The application of biotechnology to industrial production holds many promises for sustainable development, but many products still have to pass the test of economic viability. *EMBO Rep* 4:835–837
- Grogan DW (1989) Phenotypic characterization of the archaeobacterial genus *Sulfolobus*: comparison of five wild-type strains. *J Bacteriol* 171:6710–6719
- Makarova KS, Sorokin AV, Novichkov PS, Wolf YI, Koonin EV (2007) Clusters of orthologous genes for 41 archaeal genomes and implications for evolutionary genomics of archaea. *Biol Direct* 2:33
- Martusewitsch E, Sensen CW, Schleper C (2000) High spontaneous mutation rate in the hyperthermophilic archaeon *Sulfolobus solfataricus* is mediated by transposable elements. *J Bacteriol* 182:2574–2581
- Pham TK, Sierocinski P, van der Oost J, Wright PC (2009) Quantitative proteomic analysis of *Sulfolobus solfataricus* membrane proteins. *J Proteome Res* 9:1165–1172
- Redder P, She Q, Garrett RA (2001) Non-autonomous mobile elements in the crenarchaeon *Sulfolobus solfataricus*. *J Mol Biol* 306:1–6
- She Q, Peng X, Zillig W, Garrett RA (2001a) Genome evolution: gene capture in archaeal chromosomes. *Nature* 409:478
- She Q et al (2001b) The complete genome of the crenarchaeon *Sulfolobus solfataricus* P2. *Proc Natl Acad Sci USA* 98:7835–7840
- Wagner M, Berkner S, Ajon M, Driessen AJM, Lipps G, Albers SV (2009) Expanding and understanding the genetic toolbox of the hyperthermophilic genus *Sulfolobus*. *Biochem Soc Trans* 37:97–101
- Worthington P, Hoang V, Perez-Pomares F, Blum P (2003) Targeted disruption of the α -amylase gene in the hyperthermophilic archaeon *Sulfolobus solfataricus*. *J Bacteriol* 185:482–488
- Wurtzel O, Sapra R, Chen F, Zhu Y, Simmons BA, Sorek R (2010) A single-base resolution map of an archaeal transcriptome. *Genome Res* 20:133–141
- Zaparty M, Siebers B (2010) Physiology, metabolism and enzymology of thermoacidophiles. In: Horikoshi K, Antranikian G, Bull AT, Robb FT, Stetter KO (eds) *Extremophiles handbook*. Springer, Tokyo
- Zaparty M et al (2009) “Hot standards” for the thermoacidophilic archaeon *Sulfolobus solfataricus*. *Extremophiles* 14:119–142
- Zillig W, Stetter KO, Wunderl S (1980) The *Sulfolobus*-‘*Caldariella*’ group: taxonomy on the basis of the structure of DNA-dependent RNA polymerases. *Arch Mikrobiol* 125:259–269